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## Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

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### Synthesis and Studies of Modified Oligonucleotides- Directed Triple Helix Formation at the Purine-Pyrimidine Interrupted Site

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Online publication date: 09 August 2003

**To cite this Article** Jazouli, Mohamed , Guianvarc'h, Dominique , Bougrin, Khalid , Soufiaoui, Mohamed , Vierling, Pierre and Benhida, Rachid(2003) 'Synthesis and Studies of Modified Oligonucleotides- Directed Triple Helix Formation at the Purine-Pyrimidine Interrupted Site', *Nucleosides, Nucleotides and Nucleic Acids*, 22: 5, 1277 – 1280

**To link to this Article:** DOI: 10.1081/NCN-120022945

**URL:** <http://dx.doi.org/10.1081/NCN-120022945>

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## Synthesis and Studies of Modified Oligonucleotides-Directed Triple Helix Formation at the Purine-Pyrimidine Interrupted Site

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### ABSTRACT

Triple helix formation is still restricted to oligopurine-oligopyrimidine double stranded DNA target. Herein we focus on our progress achieved in nucleobase and oligonucleotide modifications area to address the chemical challenge to circumvent the recognition of a purine-pyrimidine base pair interruption in an oligopyrimidine-oligopurine DNA sequence.

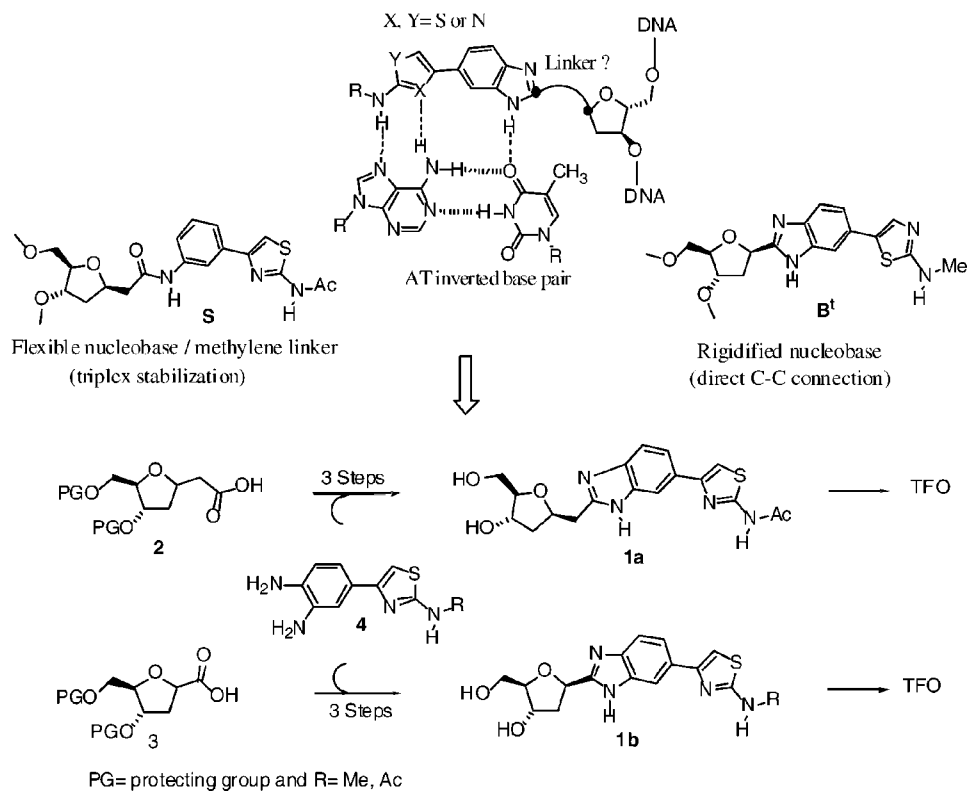
*Key Words:* Triple helix; Artificial nucleobases; AT inversion.

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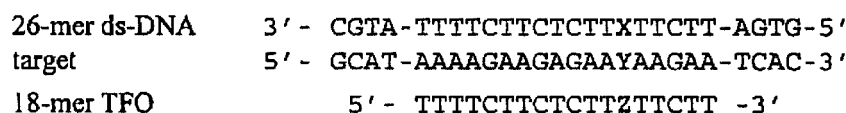


Triple helix-forming oligonucleotides bind in the major groove of oligopyrimidine-oligopurine double-stranded DNA (ds-DNA) sequences. In the pyrimidine motif, the molecular recognition process occurs between the oligonucleotide and the oligopurine strand of ds-DNA by formation of  $TA \bullet T$  and  $CG \bullet C^+$  Hoogsteen base triplets.<sup>[1]</sup> Hence, this approach (antigene strategy) provides rational basis for the development of new tools in molecular biology and for therapeutic applications.<sup>[2]</sup> Unfortunately, this strategy has a major intrinsic limitation since, the interruption of polypurine tract by one or more pyrimidine bases usually results in significant reduced triplex stability.<sup>[3]</sup> Therefore, much efforts have been undertaken to overcome this sequence limitation but, no successful results have been so far reported especially in unnatural triplexes incorporating artificial nucleobases.<sup>[4]</sup>

We have recently observed that the use of an extended heterocyclic system like **S** within TFO (**S** facing the inverted AT base pair) highly stabilized the triplex formed between this TFO and the ds-DNA target (Sch. 1). Indeed, the obtained  $T_m$  values were found to be very close to those of canonical triplexes (Table 1).<sup>[5]</sup> Moreover, the incorporation of a rigidified nucleobase **B<sup>t</sup>** into TFO, also induced an increased triplex stabilization<sup>[6]</sup> compared to the flexible analog (data not shown).



Scheme 1.



**Table 1.** Melting temperature values ( $T_m$ ) of all combinations of XYZ triplets (10 mM cacodylate, 100 mM NaCl, 10 mM MgCl<sub>2</sub>, 0.5 mM spermine, pH 6).

	Z =	T	C	G	S	Bt
XY	$T_m$ (°C)					
TA		51	31	31	42	37
CG		40	50	31	41	38
AT		33	33	45	50	43
GC		38	35	35	46	41

These findings inspired us to undertake an extensive structure-stability relationship study to assess the role of different molecular group.<sup>a</sup> Accordingly, we have recently synthesized, following two approaches, the modified nucleosides **1a** (S analog) and **1b** (B<sup>t</sup> analog) featuring (i) a rigidified thiazolyl-benzimidazole nucleobase and (ii) the common aminothiazole ring in the best configuration (nitrogen-donor instead of sulfur atom). The modified nucleoside **1b** contains, compared to **1a**, a supplementary methylene group as a spacer between the sugar and the base. Incorporation of **1a** and **1b** into TFO together with triplex hybridization studies are in progress.

### ACKNOWLEDGMENT

Financial support from CNRS-CNR (France-Maroc) is gratefully acknowledged.

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<sup>a</sup>This work on structure-stability relationship study will be published elsewhere.



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